# Species-specificity of Estradiol Regulated Growth Factors in Breast Cancer

ANDREA MANNI,\* BETTY BADGER,\* CAROL WRIGHT,\* JERRY GLENN,† S. RAFEEQ AHMED\* and LAURENCE M. DEMERS\*‡

Departments of \*Medicine, †Surgery and ‡Pathology, The Milton S. Hershey Medical Center, The Pennsylvania State University, P.O. Box 850, Hershey, PA 17033, U.S.A.

**Abstract**—Recent evidence indicates that autocrine/paracrine mechanisms may mediate the mitogenic effect of estradiol  $(E_2)$  both in human and experimental breast cancer. However, the species-specificity of  $E_2$ -regulated growth factors with regard to their biologic action has not been evaluated. To test this issue, we examined, in the soft agar clonogenic assay, the colony-stimulating activity in human breast cancers of conditioned media obtained from rat mammary carcinomas exposed to  $E_2$  (rat  $E_2$ -CM). Of 22 primary human breast cancers plated in soft agar in the absence of serum, 18 (82%) successfully grew with a mean colony number of 62.4  $\pm$  9.8 (S.E.M.) (range 14–193). Rat  $E_2$ -CM significantly stimulated colony formation in 10/18 (56%) human breast cancers to 155  $\pm$  11% (S.E.M.) of control.  $E_2$  administration (10<sup>-9</sup> M) in these tumors had a virtually identical overall effect (154  $\pm$  13% of control colony number). In the remaining eight tumors (44%), neither rat  $E_2$ -CM nor  $E_2$  had, in general, a significant colony-stimulating effect. The growth-promoting action of rat  $E_2$ -CM and  $E_2$  was not influenced by the hormone receptor status of the tumor. These results suggest that  $E_2$ -regulated growth factors may not be species-specific, at least with regard to their colony-stimulating effects in soft agar.

## INTRODUCTION

RECENT EVIDENCE indicates that autocrine/paracrine mechanisms may play an important role in supporting the growth of experimental [1, 2] and human breast cancer [3-5]. Using the hormoneresponsive N-nitrosomethyl-urea (NMU)-induced rat mammary cancer cultured in the soft agar clonogenic assay we have recently provided evidence supporting a critical role of autocrine factors in mediating hormonally stimulated growth [6-9]. We have also shown that the same experimental system is suitable to grow primary human breast cancers and to test their hormone-responsiveness [10, 11]. Thus, we felt that the soft agar clonogenic assay would be a useful tool to test the speciesspecificity of hormonally regulated growth factors with regard to their ability to stimulate mammary tumor colony formation. To address this issue, we evaluated the colony-stimulating effect of conditioned media obtained from rat mammary tumors exposed to estradiol using human breast cancers grown in soft agar in the absence of serum. The effect of exogenous administration of estradiol was also simultaneously tested in the same human breast cancer specimens. By comparing in the same tumors the colony-stimulating effect of estrogen-conditioned media with that of estradiol itself, we attempted to gain insight into the contribution of autocrine/paracrine factors to the overall growth-promoting effect of estradiol in our experimental system. Finally, the effects of these treatments on colony formation was correlated with the estrogen and progesterone receptor status of the tumor.

#### MATERIALS AND METHODS

Materials

Twenty-two human breast cancer specimens were obtained at the time of biopsy or mastectomy. Twenty were infiltrating ductal carcinomas, one was an invasive lobular carcinoma (Table 1, patient 1) and one was a medullary carcinoma (Table 1, patient 5). The mean age of the patients was 57 years (range: 27–86 years). Fifteen women were postmenopausal while seven were premenopausal. Estrogen receptor (ER) and progesterone receptor

Accepted 6 April 1988.

Address for reprints: Andrea Manni, M.D., Department of Medicine/Endocrinology, The Milton S. Hershey Medical Center, The Pennsylvania State University, P.O. Box 850, Hershey, PA 17033, U.S.A.

1350 A. Manni et al.

Table 1. Summary of the treatment effects in the 10 human breast cancers, the growth of which in soft agar was significantly stimulated by rat  $E_2$ -CM

Patient No.	Age (years)	Menopausal status			Colonies*							
			ER (fmol/mg)	PgR (fmol/mg)	Control	$E_2$ -CM		C-CM		$E_2 (10^{-9} M)$		
						No.	% of control	No.	% of control	No.	% of control	
1	68	Post	37	108	$27 \pm 0.7$	$45 \pm 0.4$	167	$28 \pm 0.7$	104	$58 \pm 2.5$	215	
2	52	Post	42	91	$59 \pm 1.2$	$74 \pm 0.7$	125	$59 \pm 1.0$	100	$72 \pm 2.1$	122	
3	60	Post	157	22	$32 \pm 1.2$	$40 \pm 1.2$	125	$32 \pm 1.2$	100	$39 \pm 0.7$	122	
4	74	Post	23	73	$193 \pm 2.4$	$244 \pm 5.9$	126	$211 \pm 8.1$	109	$239 \pm 5.2$	124	
5	30	Pre	0	0	$88 \pm 2.0$	$127 \pm 5.1$	144	$80 \pm 5.9$	91	$100 \pm 5.2 \uparrow$	114	
6	67	Post	421	57	$14 \pm 1.7$	$34 \pm 1.4$	243	$26 \pm 0.5^{+}_{+}$	186	$20 \pm 0.7$	143	
7	67	Post	0	0	$45 \pm 2.9$	$80 \pm 2.5$	178	$64 \pm 2.0$	142	$89 \pm 1.5$	198	
8	31	Pre	12	29	$63 \pm 1.5$	$97 \pm 1.3$	154	$76 \pm 3.6$	121	$95 \pm 4.2$	151	
9	75	Post	660	1210	$118 \pm 7.8$	$144 \pm 5.0$	122	$159 \pm 10.0$ §	135	$267 \pm 7.0$	226	
10	86	Post	413	79	$16 \pm 3.5$	$27\pm1.4$	169	$26 \pm 1.5$ §	163	$20 \pm 1.0 \dagger$	125	

<sup>\*</sup>Data represent actual colony number (mean  $\pm$  S.E.M. of three replicate dishes). In each of these human tumors rat E<sub>2</sub>-CM significantly stimulated colony formation over control (P < 0.05, Newman–Keul test).

(PGR) measurements were performed in all tumors. Seventeen patients were ER+ PGR+, one patient was ER- PGR+, and four patients were ER-PGR-. The rat mammary tumors used to generate our conditioned media were obtained with a single injection of NMU (Sigma Chemical Co., St. Louis, MO) (5 mg/100 g body/wt) in female Sprague-Dawley rats as previously described [12].

# Experimental design and methods

a. Preparation of the rat mammary tumor conditioned media. Single cell suspensions obtained from freshly excised NMU-induced mammary tumors were plated in soft agar in the presence or in the absence of E<sub>2</sub> (10<sup>-9</sup>M) under serum-free media conditions. The details of our culture technique, which is a slight modification of the methods described by Hamburger and Salmon [13] and Von Hoff et al. [14] have been extensively reported in recent publications [15, 16]. Conditioned media from E<sub>2</sub>treated and control dishes were obtained by adding 2 ml of Hank's balanced salt solution/dish 2 days after plating and removing it 24 h later. This method has been found to be effective in collecting secretory products of cells plated in soft agar [14, 17, 18]. E<sub>2</sub>-CM and C-CM were then concentrated on ice using a 3500 molecular weight cut-off dialysis membrane (Spectra-Por/6) with aquacide (sodium salt of carboxymethyl cellulose). The concentrated media were then treated with dextrancoated charcoal (0.25% charcoal and 0.0025% DEAE-dextran) and then kept frozen at -70°C until used in the experiments. Since the stability with time of the colony-stimulating effect of the conditioned media in our system is not known, a

new set of media was prepared on a weekly basis. Thus, a recently prepared set was always available whenever we received a human breast cancer specimen. In addition to collecting the conditioned media we verified, in each case, that E2 had exerted its known colony-stimulating effect in this system as we have previously reported [19]. Each set of conditioned media was routinely checked for the contaminating presence of estradiol measured by RIA [20]. In each case E2 levels were found to be less than 10 pg/ml. Since the samples were at least 10 times more concentrated than their final concentrations in our experiments described below, the amount of contaminating E2 was actually less than 1 pg/ml. Thus, we can reasonably exclude any contribution from contaminating E2 to the observed effects of the rat conditioned media on human mammary cancer colony formation. Estradiol (Sigma Chemical Co., St. Louis, MO) was initially dissolved in 100% ethanol and then added to both layers of the medium at the desired concentration.

b. Evaluation of the effects of the rat mammary tumor conditioned media on human breast cancer growth in soft agar. Each human tumor was plated in triplicate 35 mm Petri dishes in the absence of serum under the following experimental conditions: (1) control, no treatment, (2) rat  $E_2$ -CM, (3) rat C-CM and (4)  $E_2$   $10^{-9}$  M. Between 100,000 and 500,000 cells/dish were plated depending on the size and the cellularity of the specimens received. Obviously, each tumor was plated at the same cell density under the different experimental conditions. Immediately after plating, the dishes were inspected to exclude the presence of clumps which could erroneously be

 $<sup>\</sup>dagger P = \text{NS vs. control}$ . In the remaining eight tumors the colony-stimulating effect of  $E_2$  was significant (P < 0.05 vs. control).

 $<sup>^{\</sup>ddagger}P < 0.05$  vs. control and E<sub>2</sub>-CM.

 $<sup>\</sup>delta P < 0.05$  vs. control.

scored as colonies formed during the incubation time. We have previously consistently demonstrated the feasibility of growing experimental [16, 19] as well as human [10, 11] breast cancers under serumfree medium conditions in the soft agar clonogenic assay. The number of colonies (aggregates of more than 50 cells with a mean square diameter  $> 85 \mu m$ ) formed 6 days after plating was used to evaluate the effect of our treatments on colony formation. The validity of this parameter as an index of tumor growth has been supported by its correlation with incorporation of tritiated thymidine into DNA [21]. Simultaneously with each human tumor, an NMU rat mammary tumor was plated under identical culture conditions. This served as a positive control since we have observed that, in this system, both E<sub>2</sub> [19] and E<sub>2</sub>-CM [7] consistently stimulate colony formation. An aliquot of the same human tumor sample used for plating was also assayed for ER and PGR. The measurement of these receptors was performed according to standard techniques routinely used in our laboratory [22, 23].

#### Statistical analysis

The treatment effects on colony formation were individually evaluated for each human tumor tested by analysis of variance using the Newman-Keul test [24].

# **RESULTS**

#### Basal Growth

Eighteen of 22 (82%) primary human breast cancers successfully grew in soft agar under serum-free media conditions. The average number of colonies formed by these tumors was  $62.4 \pm 9.8$  (S.E.M.) (range: 14–193). The four tumors that did not grow were infiltrating ductal carcinomas, removed from three postmenopausal and one premenopausal women. All were estrogen and progesterone receptor positive.

Overall effects of rat mammary tumor conditioned media and estradiol on human breast cancer growth in soft agar

E<sub>2</sub>-CM significantly stimulated colony formation in 10 of 18 (56%) human breast cancers to  $155 \pm 11\%$  (S.E.M.) of control (Table 1). Rat C-CM simultaneously tested, had no effect in five human tumors (Table 1, patients 1-5) while having a significant colony-stimulating effect in the remaining five (Table 1, patients 6-10). Such colonystimulating effect was significantly less than that observed with rat E<sub>2</sub>-CM in three tumors (patients 6-8) while it was similar in the other two (patients 9 and 10). The overall colony-stimulating effect of E<sub>2</sub> in these 10 tumors was virtually identical to that of rat  $E_2$ -CM (154  $\pm$  13% of control). The effect of E2 was statistically significant in all but two tumors (Table 1, patients 5 and 10). Compared to rat E<sub>2</sub>-CM the colony-stimulating effect of E<sub>2</sub> was significantly higher (P < 0.05) in three tumors (Table 1, patients 1, 7 and 9) and significantly lower (P < 0.05) in two (Table 1, patients 5 and

In eight of 18 tumors (44%) rat  $E_2$ -CM had no colony-stimulating effect (Table 2). Administration of  $E_2$  had a remarkably similar lack of growth-promoting action. In only one of these eight tumors (Table 2, patient 6)  $E_2$  had a modest, although significant, colony-stimulating effect.

Influence of the hormone receptor status on the treatment effects

Rat  $E_2$ -CM significantly stimulated colony formation in eight of 13 (62%) ER and PGR positive human tumors and in two of five (40%) receptor negative mammary cancers. The overall effects of the rat conditioned media and  $E_2$  in the 13 receptor positive and five receptor negative tumors is illustrated in Fig. 1. It can be seen that the colony-stimulating effects of the treatments used were similar in the receptor positive and negative tumors.

Table 2. Summary of the treatment effects in the eight human breast cancers, the growth of which in soft agar was not significantly stimulated by rat E<sub>2</sub>-CM

Patient No.		Menopausal status			Colonies*							
			ER (fmol/mg)	PgR (fmol/mg)	Control	$E_2$ -CM		C-CM		$E_2 (10^{-9} M)$		
	Age (years)					No.	% of control	No.	% of control	No.	% of control	
1	71	Post	95	77	$59 \pm 4.2$	58 ± 1.4	98	$53 \pm 3.3$	90	$56 \pm 2.0$	95	
2	40	Pre	0	22	$28 \pm 2.9$	$32 \pm 2.2$	114	$28 \pm 2.9$	100	$36 \pm 3.3$	129	
3	60	Post	11	81	$62 \pm 3.1$	$70 \pm 5.0$	113	$57 \pm 3.1$	92	$65 \pm 3.0$	105	
4	71	Post	787	273	$33 \pm 1.4$	$32 \pm 1.7$	97	$31 \pm 1.2$	94	$35 \pm 3.4$	106	
5	28	Pre	24	24	$60 \pm 6.1$	$52 \pm 1.7$	87	$47 \pm 2.1$	78	$49 \pm 0.8$	82	
6	35	Pre	0	0	$88 \pm 7.0$	$86 \pm 7.0$	98	$75 \pm 1.4$	85	$107 \pm 2.8 \dagger$	122	
7	64	Post	39	18	$83 \pm 3.1$	$82 \pm 2.1$	99	$78 \pm 3.5$	94	$73 \pm 1.8$	88	
8	27	Pre	0	0	$56 \pm 2.2$	$55 \pm 1.5$	98	$56 \pm 2.9$	100	$57 \pm 2.9$	102	

<sup>\*</sup>Data are expressed as in Table 1.

 $<sup>\</sup>dagger P < 0.05$  vs. control and E<sub>2</sub>-CM. In the remaining seven tumors E<sub>2</sub> had no significant effect on colony formation.

1352 A. Manni et al.

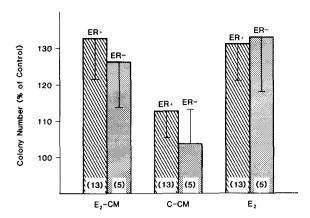


Fig. 1. Influence of the estrogen receptor status on the effects of the rat conditioned media and  $E_2$  on human breast cancer growth in soft agar. Data represent means  $\pm$  S.E.M. The number of human tumors tested is shown in parentheses. The 13 ER+ tumors were also PGR+. Of the five ER- tumors, four were also PGR- while one was low positive (22 fmol/mg cytosol protein).

## **DISCUSSION**

Despite considerable evidence suggesting a major role of autocrine/paracrine factors as mediators of human and experimental breast cancer growth [1-5], the issue of species-specificity with regard to the biological activity of hormonally regulated growth factors has not been previously addressed. The availability of an in vitro culture system which allows the growth of both primary human and experimental mammary tumors under serum-free media conditions provided us with the opportunity to approach this question. In previous experiments we had shown that conditioned media obtained from NMU rat mammary tumors exposed to estradiol [7] and prolactin [8, 9] were able to stimulate the growth of this experimental mammary tumor model in the soft agar clonogenic assay. Those results provided support for the contention that autocrine/ paracrine factors are important mediators of hormonally induced tumor growth in our system. The present experiments were designed to test whether conditioned media obtained from estradiol-exposed rat mammary tumors would be able to stimulate the growth of human breast cancers thus testing whether hormonally regulated growth factors are species-specific at least with regard to their colonystimulating effect in soft agar. Our data provide evidence for a lack of species-specificity in this regard since a significant fraction of human breast cancers (56%) was stimulated to grow in soft agar by rat E2-CM. It is conceivable that in this as well as our previous reports [7-9], we may actually have underestimated the biologic activity of the conditioned media since treatment of the media with dextran-coated charcoal may have partially removed the growth factors. On the other hand, such treatment is necessary in order to remove the

contaminating presence of even small amounts of estradiol which would confound the interpretation of our results. Our biologic data are in agreement with the available information on the biochemical characterization of growth factors produced by experimental and human breast cancers. Zwiebel et al. [1] detected high molecular weight TGFα-like growth factors in acid ethanol extracts of NMUinduced rat mammary tumors. These authors found similar growth factors in tumor extracts and conditioned media obtained from another hormoneresponsive experimental breast cancer, i.e. the 7,12dimethyl-benza(a)anthracene-induced rat mammary tumor [1]. Recently, high molecular weight TGFα-like growth factors have been found to be secreted by a variety of human breast cancer cell lines [25]. Furthermore, in the hormone-responsive cell lines  $TGF\alpha$ -like activity has been found to be highly estrogen inducible [25]. Taken together, these data indicate that hormonally regulated growth factors, potentially involved in the growth of hormone-responsive human and experimental mammary tumors, may be similar in nature.

The correlation between the colony-stimulating effect, or lack of it, of E2-CM and E2 observed in our experiments (Tables 1 and 2) deserves some comment. Although autocrine/paracrine factors are considered to be important mediators of estradiol effects on breast cancer cell proliferation, the relative contribution of these 'distal messages' to the overall estrogen effect is not fully established. Our experiments testing simultaneously the colony-stimulating effects of E2 and E2-CM partially address this issue. The similarity of the growth-promoting action of these two treatments indicate that, in our experimental system, autocrine/paracrine factors play a predominant role in estradiol-stimulated growth of hormone-responsive breast cancer. If E2, in fact, would promote tumor growth predominantly through alternative mechanisms, one would anticipate that a significant fraction of tumors not stimulated by E2-CM (i.e. the E2-regulated growth factors) would be, instead, stimulated to grow by E2 itself. However, this was not observed in our experiments as shown in Table 2, where E2 essentially failed to stimulate colony formation in tumors that were unresponsive to E2-CM. Our data also do not support the concept that hormone-independent breast cancer cells are stimulated by growth factors produced by hormone-responsive cells under estradiol influence. If this were the case, one would anticipate that a significant fraction of tumors that are insensitive to E2 would be stimulated instead by E<sub>2</sub>-CM (i.e. estradiol-regulated growth factors). This, however, again was not observed since tumors which were insensitive to E2 administration were, in general, also similarly insensitive to E<sub>2</sub>-CM.

Finally, we observed that the colony-stimulating effects of the E<sub>2</sub>-CM and E<sub>2</sub> were not influenced by

the hormone receptor status of the tumor. This finding is in agreement with our previous observation that a significant fraction of estrogen receptor negative human breast cancers are stimulated to grow in soft agar by the addition of estradiol [10] and also prolactin [11]. Thus, it appears that tumors defined as receptor negative on the basis of the biochemical assay probably contain clones of hormone-responsive cells as recently indicated by assays that employ immunohistochemical methods

[26]. Our findings, in this regard, are also in agreement with the recent observation that administration of estrogens to women with breast cancer as a means of synchronizing tumor growth has been found to stimulate DNA synthesis both in receptor positive and receptor negative tumors [27, 28].

Acknowledgements—The authors wish to thank Mrs. Sandra Christian for her excellent secretarial assistance. This paper is supported in part by Program Project Grant No. P01-CA40011.

## REFERENCES

- 1. Zwiebel JA, Davix MR, Kohn E et al. Anchorage-independent growth-conferring factor production by rat mammary tumor cells. Cancer Res 1982, 42, 5117-5125.
- Hiragun A, Yoshida Y, Sato M et al. Isolation of two syngeneic cell lines from a rat mammary carcinoma: growth factor production by neoplastic epithelial cells. J Natl Cancer Inst 1985, 75, 471-482.
- Chalbos DF, Vignon F, Keydar I et al. Estrogens stimulate cell proliferation and induce secretory proteins in a human breast cancer cell line (T<sup>47</sup>D). J Clin Endocrinol Metab 1982, 55, 276–283.
- Dickson RB, McManaway ME, Lippman ME. Estrogen-induced factors of breast cancer cells partially replace estrogen to promote tumor growth. Science 1986, 232, 1540–1453.
- 5. Huff KK, Kauffman D, Gabbay KH et al. Secretion of an insulin-like growth factor-I-related protein by human breast cancer cells. Cancer Res 1986, 46, 4613-4619.
- Manni A, Wright C, Luk GD et al. Polyamines and the synthesis of estradiol-regulated growth factors in rat mammary cancer in culture. Breast Cancer Res Treat 1987, 9, 45-51.
- 7. Manni A, Wright C, Feil P et al. Autocrine stimulation by estradiol-regulated growth factors of rat hormone-responsive mammary cancer: interaction with the polyamine pathway. Cancer Res 1986, 46, 1594-1598.
- 8. Manni A, Pontari M, Wright C. Autocrine stimulation by prolactin of hormone-responsive breast cancer growth in culture. *Endocrinology* 1986, 117, 2040–2043.
- Manni A, Wright C, Hsu C-J et al. Polyamines and autocrine control of tumor growth by prolactin in experimental breast cancer in culture. Endocrinology 1986, 119, 2033–2037.
- 10. Manni A, Wright C, Pontari M et al. Hormone dependency of human breast neoplasms culture in vitro in the stem cell assay. J Natl Cancer Inst 1985, 74, 767-770.
- 11. Manni A, Wright C, Davis G et al. Promotion by prolactin of the growth of human breast neoplasms cultured in vitro in the soft agar clonogenic assay. Cancer Res 1986, 46, 1669–1672.
- 12. Arafah BM, Finegan HM, Roe J et al. Hormone dependency in N-nitrosomethylurea-induced rat mammary tumors. Endocrinology 1982, 11, 584-588.
- 13. Hamburger AW, Salmon SE. Primary bioassay of human myeloma stem cells. *J Clin Invest* 1977, **60**, 846–854.
- Von Hoff DD, Casper J, Bradley E et al. Direct cloning of neuroblastoma cells in soft agar culture. Cancer Res 1980, 40, 3591-3597.
- Manni A, Wright C. Effect of tamoxifen and α-difluoromethyl-ornithine on clones of nitrosomethylurea-induced rat mammary tumor cells grown in soft agar. Cancer Res 1983, 43, 1084–1088.
- 16. Manni A, Wright C. Polyamines as mediators of the effect of prolactin and growth hormone on the growth of N-nitrosomethylurea-induced rat mammary tumor cultured *in vitro* in soft agar. J Natl Cancer Inst 1985, **74**, 941–944.
- Arafah BM, Wilhite BL, Rainieri J et al. Inhibitory action of bromocriptine and tamoxifen on the growth of human pituitary tumors in soft agar. J Clin Endocrinol Metab 1983, 57, 986-992.
- 18. Bradley EC, Reichert CM, Brennan MF et al. Direct cloning of human parathyroid hyperplasia cells in soft agar culture. Cancer Res 1980, 40, 3694-3696.
- 19. Manni A, Wright C. Polyamines as mediators of estrogen action on the growth of experimental breast cancer in rats. J Natl Cancer Inst 1984, 73, 511-514.
- 20. Haning RV, Meier SM, Boehnlein LM et al. Two direct radioimmunoassays for 17α-estradiol evaluation for use in monitoring in vitro fertilization. Clin Chem 1984, 30, 787-790.
- 21. Manni A, Wright C. Assessment of mitogenesis of the hormone-responsive N-nitrosomethyl-urea-induced rat mammary tumor grown in culture in soft agar using [3H]thymidine incorporation into DNA. Breast Cancer Res Treat 1983, 3, 287-292.
- 22. Feil PD, Klase JC, Margets MJ. Use of medroxyprogesterone acetate to measure cytosol progestin receptors in human breast cancer. In: Program of the 61st Annual Meeting of the American Endocrine Society, Anaheim, CA, 13–15 June 1979, Abstract 310.
- 23. McGuire WL, Dela Garza M, Chamness GC. Evaluation of estrogen receptor assays in human breast cancer tissue. *Cancer Res* 1977, 37, 637-639, 1977.

- 24. Winer BJ. Statistical Principles in Experimental Design. New York, McGraw-Hill, 1971, 185-196.
- 25. Dickson RB, Bates SE, McManaway ME et al. Characterization of estrogen responsive transforming activity in human breast cancer cell lines. Cancer Res 1986, 46, 1707-1713.
- 26. Marchetti E, Querzoli P, Moncharmont B et al. Immunocytochemical demonstration of estrogen receptors by monoclonal antibodies in human breast cancer: correlation with estrogen receptor assay by dextran-coated charcoal method. Cancer Res 1987, 47, 2508–2513.
- 27. Dao TL, Sinha DK, Nemoto T, Patel J. Effect of estrogen and progesterone on cellular replication of human breast tumors. *Cancer Res* 1982, **42**, 359–362.
- 28. Conte PF, Frashini G, Alama A et al. Chemotherapy following estrogen-induced expansion of the growth fraction of human breast cancer. Cancer Res 1985, 45, 5926-5930.